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# UNVEILING GENETIC VARIABILITY OF PHYSIOLOGICAL TRAITS DRIVING DROUGHT TOLERANCE IN CHICKPEA (CICER ARIETINUM L.)

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ABSTRACT

Chickpea (Cicer arietinum L.) is one of the important legumes widely grown for dietary proteins in semi-arid Mediterranean climatic conditions. Its crop yield is highly susceptible to abiotic stresses such as heat, frost and drought, especially during the reproductive phase, causing major yield losses and production instability worldwide. With climate change intensifying the frequency and severity of drought and heat stress, the development of stress-tolerant varieties is crucial for sustaining and enhancing chickpea production. Many researchers have emphasized the genetic improvement of yieldrelated traits under drought conditions as the primary avenue for developing high-yielding, droughttolerant varieties. However, drought tolerance can also be achieved through the selection of physiological traits directly associated with stress adaptation. Therefore, an investigation was conducted at the Regional Agricultural Research Station, Vijayapur, during the post-rainy season to assess the genetic differences for physiological traits among chickpea genotypes. The experimental material consisted of 30 chickpea genotypes including six checks. Analysis of variance revealed significant genotypic differences for all the four physiological characters with wide range of variations. Genotypic and phenotypic co-efficient of variation were high for proline content under both stress and non-stress condition. Broad sense heritability was higher for all the four traits. High heritability coupled with high genetic advance over mean was exhibited by SPAD chlorophyll meter reading and Proline content at 45 days after sowing under both stress and non-stress conditions. Based on drought indices, DBGV211, DIBG205, DBGV219, DBGV210, ICCV191116 and ICCV191102 were identified as superior genotypes and can be used as parents to develop breeding populations to identify genomic regions linked to specific traits.

Keywords: Broad sense heritability, Chickpea, SPAD chlorophyll meter reading, Proline content

## Introduction

Chickpea is the most important legume crop and a source of nutrition to millions of people globally due to its richness in protein, fibre and minerals. It also remediates the soil by its ability to fix nitrogen in a symbiotic relationship with rhizobacteria upon nodulation. It belongs to the family Fabaceae, subfamily Faboideae. The genus *Cicer* consists of 44 species including 35 perennial and 8 wild species, with *Cicer arietinum* L. as the domesticated species. The crop is self-pollinated, diploid (2n=2x=16) with a

relatively small genome (740 Mb). The cultivated chickpea is divided into two groups as microsperma and macrosperma on the basis of plant and seed characteristics (Varshney *et al.*, 2014).

Terminal drought is major limiting factor because in majority of the areas, chickpea is cultivated as post rainy season crop. The crop is generally planted after the main rainy season and grown on stored soil moisture, making terminal drought stress a primary constraint to productivity (Leport *et al.*, 1999 and Serraj *et al.*, 2004). The crop experiences drought

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stress from late vegetative stages until maturity. Drought stress intensity varies from year to year and from place to place, depending on amount and distribution of rainfall, as well as spring and early summer temperatures. Limited irrigation to meet crop needs at critical times of growth and development may be vital for realizing chickpea cultivars' yield potential. However, the majority of chickpea production takes place in areas where irrigation is not available. Farmers with limited resources who grow chickpeas find it difficult to provide supplemental irrigation. Therefore, developing high yielding drought tolerant genotypes is the only way to address or overcome this constraint.

JG 11 and JAKI 9218 are widely cultivated chickpea varieties in North Karnataka; however, their performance is largely confined to the normal sowing window (September 25th-October 30th). When sown beyond this period, these varieties experience substantial yield losses, primarily due to exposure to high-temperature stress during terminal reproductive phase, which adversely affects flowering, pod set and seed filling (Devasirvatham et al., 2015). Availability of irrigation water in command areas also necessitates the need to identify suitable breeding lines for late sown cultivation. In this context, the well-adapted, drought-tolerant identification of genotypes is crucial for improving both the production and productivity of chickpea.

Understanding the variability in physiological traits is essential for improving drought tolerance in crops. Traits such as chlorophyll content, membrane relative water content, stability, and proline accumulation reflect how plants cope with water stress. When considerable genetic variability exists for these traits, it gives plant breeders a fortuitous to select genotypes that perform better under (Kashiwagi et al., 2005). Practicing selection for these traits in breeding programs helps in developing crop varieties that are more resilient and stable in yield even during dry conditions. This ultimately supports sustainable crop production and food security in drought-prone areas. The present investigation was aimed to estimate the genetic variability, heritability and genetic advance for different physiological traits along with drought tolerance indices based on yield performance.

#### **Materials and Methods**

The experimental material comprised 30 chickpea genotypes, including six standard checks; A-1, SA-1, ICC4958, BGD111-1, JAKI9218 and JG11. The trial was conducted under both stress (Rainfed) and non-stress (Irrigated) conditions during the *Rabi* 2021-22

season at the rainout shelter, III Block, Regional Agricultural Research Station (RARS), College of Agriculture, Vijayapur. A Randomized Block Design (RBD) with two replications was employed to ensure reliable evaluation of genotypic performance. The spacing was maintained at 30 cm between rows and 10 cm between plants, with each plot measuring 36 m<sup>2</sup> (2  $m \times 18$  m). The crop was raised with all standard agronomical package of practices i.e. fertilizer dose N, P and K applied as basal and all plant protection measures were adopted to raise healthy crop. The observations were recorded on SPAD chlorophyll meter reading (SCMR), membrane stability index (MSI), relative water content (RWC) and proline content (PC) at 45 and 75 days after sowing. The various genetic parameter viz. genotypic and phenotypic coefficients of variation, broad sense heritability, genetic advance over mean were formulated as suggested by Burton and Devane (1952).

#### Statistical analysis

The mean data of physiological parameters recorded for all the genotypes, were subjected to various standard statistical procedures to estimate mean, range, ANOVA and variability parameters. The data was analyzed using WINDOSTAT ver. 9.1 software programme. Genotypic and phenotypic coefficients of variability were computed as per the formula. Broad sense heritability was calculated as the ratio of genotypic variance to the total phenotypic variance, as suggested by Hanson et al. (1956) and expressed as percentage. Genetic advance and Genetic advance over mean were estimated by using the formula given by Johnson et al. (1955). In addition, five drought tolerance indices include Drought and Productivity Index (DPI), Drought tolerance efficiency productivity (DTE), Mean (MP), Drought susceptibility index (DSI) and Tolerance index (TOL) were estimated using the following formula.

- Drought and Productivity Index (DPI) =  $(3Y_S+1Y_{NS}-2DC_S)/2$
- Drought susceptibility index (DSI) = 1-(Y<sub>S</sub> /Y<sub>NS</sub>) / D (Fischer and Maurer, 1978)
- Mean productivity (MP) = (Y<sub>NS</sub> +Ys) / 2 (Rosielle and Hamblin, 1981)
- Tolerance to Drought index (TDS) =  $Y_{NS} Y_{S}$
- Drought tolerance efficiency (DTE) =  $(Y_S / Y_{NS}) X$ 100

### Where

Y<sub>S</sub> = Grain yield of the genotype under moisture stress condition

 $Y_{NS}$  = Grain yield of the genotype under non-stress condition

DC<sub>S</sub> = Grain yield of drought tolerant check under stress condition

Mean grain yield of all strain under  $D = 1 - \frac{\text{moisture stress condition}}{\text{Mean grain yield of all strains under non}}$ - stress condition

#### **Results and Discussion**

The mean performance of genotypes showed significant variation for all four physiological traits under both stress and non-stress conditions, indicating adequate scope for effective selection in crop improvement (Fig. 1). The analysis of variance carried out on four physiological traits reveal that mean sum of squares due to genotypes was significant at both 45 and 75 days after sowing under stress and non-stress conditions, indicating the presence of substantial genetic variability for the traits studied among the genotypes (Table 1). Variability within a population is fundamental to crop improvement, as it provides the basis for identifying and selecting superior genotypes in a breeding programme. It can be quantified through several approaches, among which the phenotypic coefficient of variation (PCV) and genotypic coefficient of variation (GCV) are the most widely used indicators. Generally, higher PCV values compared to GCV suggest a positive influence of the environment on the expression of physiological traits. Estimates of PCV, GCV, heritability, and genetic advance over mean for physiological traits at both intervals under both stress and non-stress conditions are presented in Table 2.

The phenotypic and genotypic coefficient of variation were high for PC at 45 days after sowing under both stress and non-stress condition, indicates ample improvement in the trait when selection was practiced. However, the magnitude of difference between GCV and PCV was very low representing that they were majorly under genetic control and the influence of environment was negligible. These results were in conformity with earlier reports of Singh et al. (2013) and Singh et al. (2021). Moderate GCV and PCV were observed for PC at 75 days, SCMR at 45 days, MSI at both intervals and RWC at 75 days under both conditions, indicates the moderate variability in these traits. Higher estimates of PCV than that of the corresponding GCV indicates the positive influence of environmental factors in expression of these traits and were in line with the findings of Meena et al. (2006), Singh et al. (2013) and Manasa et al. (2020). Low PCV

and GCV for RWC at 45 days under both conditions indicated limited genetic variation, making selection less effective for this trait.

The proportion of variability due to genetic cause is projected as heritability. Heritability imprints on the transmission of traits from parents to offspring. It plays a key role in the selection process in plant breeding as it is estimated from fixable genetic variance. High heritability was recorded for SCMR at 45 days, MSI at 75 days, and PC at 45 days under both conditions, suggesting genetic improvement of these traits will be effective. Although high heritability indicates the reliability of phenotypic performance in selection, it does not always reflect the expected genetic gain.

High heritability accompanied with high genetic advance over mean were observed for SCMR and PC at 45 days under both stress and non-stress conditions, MSI at both intervals, and PC at 75 days under nonstress conditions, indicates that the majority of the variations in this character are attributable to additive gene effects and selection for these traits may be effective. High heritability coupled with moderate genetic advance were exhibited by SCMR and RWC at 75 days under both conditions, MSI at both intervals, RWC at 45 days, and PC at 75 days under stress condition indicate a combination of additive and nonadditive gene action. High heritability with low genetic advance exhibited by RWC at 45 days, suggested the prevalence of non-additive gene action. These findings corroborate earlier reports by Singh et al. (2021), Sanjay et al. (2019), and Gautam et al. (2021).

Based on yield performance under stress and nonstress conditions, the genotypes were screened using drought tolerance indices like TDS, MP, DSI, TDE and DPI. The genotypes DBGV211, DIBG205, DBGV219, DBGV210, ICCV191116, and ICCV191102 performed well under both stress and non-stress conditions, indicating their potential for use in future breeding programmes without any yield penalty (Table 3).

#### Conclusion

Genetic variability study revealed high phenotypic and genotypic coefficient of variation, heritability and genetic advance over mean for traits like membrane stability index and proline content indicate the predominance of additive gene effect and therefore simple selection based on phenotypic performance is likely to yield beneficial results in improving these characters will help in development of drought tolerant high yielding superior genotypes. The genotypes performed superior under both stress and non-stress condition can be used as parents in crossing program to

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develop breeding populations without compromising yield trait to identify genes/QTLs governing the specific trait and further can also be used in marker

assisted selection for developing drought resilient genotypes.

**Table 1:** Analysis of variance for physiological related traits in chickpea genotypes under stress and non-stress conditions

| Source of variations | Situation | DF | SCMR    | SCMR    | RWC     | RWC     | MSI      | MSI        | PC                 | PC     |
|----------------------|-----------|----|---------|---------|---------|---------|----------|------------|--------------------|--------|
|                      |           |    | @45     | @75     | @45     | @75     | @45      | <b>@75</b> | @45                | @75    |
| Replications         | NS        | 1  | 0.02    | 0.75    | 95.63   | 99.61   | 4.68     | 25.29      | 1.19               | 1.97   |
|                      | S         | 1  | 4.89    | 0.75    | 59.28   | 79.61   | 61.49    | 15.29      | 12.48              | 4.53   |
| Genotypes            | NS        | 29 | 49.66** | 32.47** | 87.33** | 116.40* | 244.45** | 112.96*    | 9.74**             | 3.91** |
|                      | S         | 29 | 39.02** | 32.47** | 73.96** | 116.39* | 140.10** | 102.96**   | 18.33 <sup>*</sup> | 8.03** |
| Residual             | NS        | 29 | 7.12    | 10.37   | 46.71   | 38.08   | 13.07    | 15.11      | 2.29               | 0.47   |
|                      | S         | 29 | 3.22    | 10.37   | 20.71   | 38.08   | 47.97    | 15.11      | 4.61               | 2.23   |
| CD @ 5%              | NS        | -  | 5.46    | 6.59    | 13.98   | 12.62   | 7.40     | 7.95       | 3.10               | 1.40   |
|                      | S         | -  | 3.67    | 5.29    | 9.31    | 10.62   | 14.17    | 5.95       | 4.39               | 3.06   |
| CV%                  | NS        | -  | 7.41    | 9.42    | 8.77    | 9.62    | 5.55     | 6.17       | 18.08              | 7.64   |
|                      | S         | -  | 5.17    | 8.32    | 5.94    | 7.62    | 10.14    | 4.17       | 14.68              | 10.62  |
| S.E. d               | NS        | -  | 2.67    | 3.22    | 6.84    | 6.17    | 3.62     | 3.89       | 1.52               | 0.69   |
|                      | S         | -  | 1.79    | 1.22    | 4.55    | 4.17    | 6.92     | 3.88       | 2.14               | 1.49   |

<sup>\*</sup>Significant at P = 0.05, \*\* P = 0.01.

SCMR-SPAD chlorophyll meter reading; RWC- relative water content; MSI-Membrane stability index; PC-Proline content

**Table 2:** Estimation of genetic variability parameters for physiological related traits in chickpea genotypes under stress and non-stress condition

| Particulars                  | T  | raits | SCMR<br>@45 | SCMR<br>@75 | RWC<br>@45 | RWC<br>@75 | MSI<br>@45 | MSI<br>@75 | PC<br>@45 | PC<br>@75 |
|------------------------------|----|-------|-------------|-------------|------------|------------|------------|------------|-----------|-----------|
| GCV                          | NS |       | 12.81       | 8.43        | 5.78       | 12.98      | 16.51      | 12.31      | 23.01     | 14.6      |
|                              | S  |       | 12.18       | 9.72        | 6.74       | 9.76       | 9.94       | 11.1       | 20.21     | 12.08     |
| PCV                          | NS |       | 14.8        | 15.4        | 10.51      | 14.21      | 17.42      | 15.3       | 29.27     | 16.48     |
|                              | S  |       | 13.23       | 13.53       | 8.98       | 13.7       | 14.2       | 12.7       | 23.15     | 16.08     |
| MEAN                         | NS |       | 36.01       | 34.21       | 77.92      | 64.13      | 65.15      | 74.5       | 8.38      | 8.98      |
|                              | S  |       | 34.73       | 34.2        | 76.6       | 64.13      | 68.27      | 63.02      | 14.63     | 14.09     |
| GA                           | NS |       | 8.22        | 6.45        | 5.11       | 10.65      | 21         | 14.65      | 3.13      | 2.39      |
|                              | S  |       | 8.02        | 4.92        | 7.97       | 9.18       | 9.79       | 12.59      | 4.17      | 2.64      |
| h <sup>2</sup> <sub>bs</sub> | ]  | NS    | 74.89       | 51.57       | 62.39      | 65.70      | 89.84      | 76.40      | 61.83     | 78.5      |
|                              | S  |       | 84.73       | 62.27       | 56.24      | 60.07      | 48.99      | 56.40      | 60.79     | 56.43     |
| GAM (%)                      | ]  | NS    | 22.83       | 17.76       | 6.56       | 16.98      | 32.24      | 22.76      | 37.28     | 26.65     |
|                              | S  |       | 23.09       | 14.38       | 10.41      | 14.31      | 14.33      | 19.98      | 28.51     | 18.7      |
| RANGE                        | NS | Min   | 25.5        | 27.09       | 55.59      | 54.31      | 44.5       | 47.9       | 3.19      | 7.17      |
|                              |    | Max   | 45.37       | 41.72       | 92.5       | .87.00     | 85.2       | 83.7       | 11.96     | 13.91     |
|                              | NS | Min   | 26.3        | 23.09       | 58.41      | 44.31      | 42.17      | 43.5       | 10.23     | 6.81      |
|                              |    | Max   | 45.3        | 36.02       | 87.58      | 88.87      | 83.57      | 72.94      | 22.72     | 18.32     |

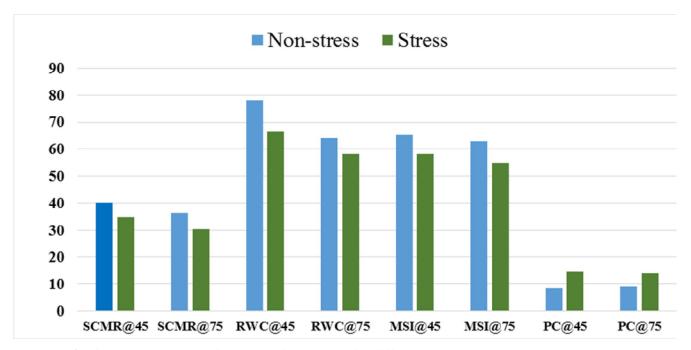
Standard range= (maximum – minimum) / mean

SCMR-SPAD chlorophyll meter reading; RWC- relative water content; MSI-Membrane stability index; PC-proline content

**Table 3:** Top six potential genotypes based on drought tolerance indices

| ENTRIES    | PYIR     | PYS     | TDS    | MP       | DSI     | DTE     | DPI     |
|------------|----------|---------|--------|----------|---------|---------|---------|
| DBGV211    | 118.931  | 106.75  | 21.655 | 117.58   | -15.46  | 147.58  | 168.755 |
| DIBG205    | 206.9262 | 148.5   | 97.425 | 158.215  | -0.77   | 91.775  | 207.015 |
| DBGV219    | 151.165  | 112.47  | 38.695 | 131.8175 | -62.315 | 130.64  | 236.275 |
| DBGV210    | 194.0117 | 138.255 | 55.755 | 166.1325 | -1.985  | 104.505 | 319.895 |
| ICCV191116 | 193.8485 | 139.1   | 94.75  | 141.475  | -1.315  | 78.23   | 223.95  |
| ICCV191102 | 244.8    | 191.81  | 67.01  | 203.305  | -13.785 | 144.255 | 236.93  |

**PYIR**=Irrigated yield; **PYRF**=Rain fed yield; **TDS**=Tolerance to drought stress; **MP**=Mean productivity; **DSI**=Drought susceptibility index; **DTE**=Drought tolerance efficiency; **DPI**=Drought and productivity index



**Fig. 1 :** Overall mean performance of genotypes for different physiological traits at two intervals under both stress and non-stress conditions

#### References

Burton, G.W. and Devane, E.M. (1952). Estimating heritability in tall fescue (*Festuca circunclinaceae*) from replicated clonal material. *Agron. J.*, **45**, 478-481.

Fischer, R.A. and Maurer, R. (1978). Drought resistance in spring wheat cultivars, I. Grain yield response. *Aust. J. Agric. Res.*, **29**, 897-912.

Devasirvatham, V., Gaur, P.M., Raju, T.N., Trethowan, R.M. and Tan, D.K.Y. (2015). Field response of chickpea (Cicer arietinum L.) to high temperature. *Field Crops Res.*, **172**, 59-71.

Fischer, R.A. and Maurer, R. (1978). Drought resistance in spring wheat cultivars, I. Grain yield response. *Aust. J. Agric. Res.*, **29**, 897-912.

Gautam, A., Panwar, R.K., Verma, S.K., Arora, A., Gaur, A.K. and Chauhan, C. (2021). Assessment of genetic variability parameters for yield and its components in chickpea (*Cicer arietinum L.*). *BFAIJ*, **13**(2), 651-655.

Hanson, C.H., Robinson, H.F. and Comstock, R.K. (1956). Biometrical studies on yield in segregating population of Korean Lasphadezia. Agron. J., 48, 314-318.

Johnson, H.W., Robinson, H.F. and Comstock, R.E. (1955). Estimation of genetics and environmental variability in soybean. Agron. J., 47, 314-318.

Kashiwagi, J., Krishnamurthy, L., Upadhyaya, H.D., Krishna, H., Chandra, S., Vadez, V. and Serraj, R. (2005). Genetic variability of drought-avoidance root traits in the minicore germplasm collection of chickpea (Cicer arietinum L.). *Euphytica*, **146**(3), 213-222.

Leport, L., Turner, N.C., French, R.J., Barr, M.D., Duda, R., Davies, S.L., Tennant, D. and Siddique, K.H.M. (1999). Physiological responses of chickpea genotypes to terminal drought in a Mediterranean-type environment. *Eur. J. Agron.*, **11**, 279-291.

Manasa, B., Shanthi Priya, Jayalaxmi, V. and Umamaheshwari, P. (2020). Genetic variability studies in extra-large and large seeded Kabuli chickpea (*Cicer arietinum L.*). *Elect. J. Pl. Breed.*, **11**, 2-8.

Meena, H.S., Kumar, J.S. and Deshmukh, P.S. (2006). Genetic variability studies for traits related to drought tolerance in chickpea. *Indian J. Genet.*, **66**(2), 140.

Rosielle, A.A. and Hamblin, J. (1981). Theoretical aspects of selection for yield in stress and non-stress environments. *Crop Sci.*, 21, 943-946.

Sanjay, K., Suresh, B.G., Anand Kumar and Lavanya, G.R. (2019). Genetic variability, correlation and path coefficient analysis in chickpea (*Cicer arietinum* L.) for yield and its component traits. *Int. J. Curr. Microbiol. App. Sci.*, 8(12), 2341-2352.

Serraj, R.L., Krishnamurthy, J., Kashiwagi, J.M., Kumar, J.S., Jundra and Crouch, J.H. (2004). Variation in root traits of chickpea (*Cicer arietinum* L.) grown under terminal drought. *Field Crops Res.*, **88**(2/3), 115-127.

Singh, A.K. and Singh, A.P. (2013). Study of genetic variability and interaction of some quantitative traits in chickpea (*Cicer arietinum L.*). *J. Multidiscip. Adv. Res.*, **2**(2), 87-94.

Singh, B. and Mishra, V.K.S. (2021). Genetic variability, path analysis and relationship among quantitative traits in chickpea (*Cicer arietinum* L.) genotypes. *Field Crops Res.*, **213**, 54-67.

Varshney, R.K., Thudi, M., Nayak, S.N., Gaur, P.M., Kashiwagi, J., Krishnamurthy, L., Jaganathan, D., Koppolu, J., Bohra, A., Tripathi, S. and Rathore, A. (2014). Genetic dissection of drought tolerance in chickpea (*Cicer arietinum* L.). *Theor. Appl. Genet.*, 127(2), 445-462.